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### 13. SUPPLEMENTARY NOTES

### 14. ABSTRACT

This project aims to determine the role of tumor stroma in prostate cancer biology. To do this, we are using a model of human embryonic stem cell (hESC) differentiation that was established in our laboratory. Using hESC-derived prostatic epithelial cells, we will test whether or not tumor stroma derived from human prostate cancer specimens will induce and initiate carcinogenesis.

Our first task has been to optimize our current protocols of hESC differentiation into prostate. Ideally, we will eliminate the small percentage of hESCs that spontaneously differentiate into non-prostatic structures in tissue grafts in order to work with a pure population of prostatic cells. Work towards this aim is in progress. In the following 6-12 months, we will begin to isolate prostatic stem cells from our hESC-derived tissues, and subsequently initiate experimental studies with human cancer stroma enriched cell populations.

### 15. SUBJECT TERMS

None provided.

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### Introduction

The role of tum or stroma in prostate cancer biology is equivocal. Current dogm a suggests that prostate carcinogenesis is a multi-step process involving genetic alterations in the epithelium that drives the progressive transformation of normal human cells into highly malignant derivatives. It is evident that tumor stroma is able to promote progression of tumorigenesis, but whether it also plays a critical role in the initiation of tumor formation is unclear.

Epithelial cells are under the control of the underlying mesenchymal cells during embryogenesis and throughout life; it is therefore our **hypothesis that the prostatic stromal cells have the capacity to initiate carcinogenesis in normal epithelial cells**. In order to address the issue of tumor initiation, we propose to use norm al human prostate epithelium generated f rom human embryonic stem cells in tissue re combination studies with tum or stroma from human prostate cancer patients.

In this project, we propose to use hum an embryonic stem cells as a source of norm al human prostate epithelial cells. Normal human prostate tissue from adult men in the prime of his life is difficult to obtain, and human fetal tissue is of limited availability. We successfully achieved this goal and published the findings in Nature Methods (Taylor, et al., 2006).

### **Body**

Although funds for this project were released in the first half of 2008, we spent several months getting ethics approval for both animal and human experimental procedures. Although existing institutional approvals were in place, we were required to make several amendments to meet the requirements of US Department of Defense legislation and regulations. This was successfully completed and we obtained formal approval from Human Research Protection Office (Office of Research Protections, U.S. Army Medical Research and Materiel Command) on 7<sup>th</sup> January 2009 and USAMRMC Animal Care & Use Review Office on 31<sup>st</sup> December 2008.

During 2008, we employed staff on this project, although they were restricted in the experimental procedures they were able to perform due to pending ethics approval. Therefore, work on this project has progressed slowly, although we have established several key experimental techniques to date, including work towards Task 1 as follows:

# Task 1 (Aim 1.1): To improve our current method of directing hESC differentiation to obtain genetically normal human prostatic epithelial cells [Years 0-1.5].

- a. <u>Culture and m aintenance of human e mbryonic stem cells (hESCs); including routine karyotyping and identification of other pluripotent markers of undifferentiated hESCs.</u>
  - Culture of hESCs has been initiated and maintained. We currently have up to 3 hESC lines growing in the lab oratory for use in th is project. These cell lines are rou tinely passaged and over a period of tim e, have be en proven to m aintain pluripotentiality and stable karyotype profiles. Staff are well trained in culture of hESCs which is vital to the success of this project.
- b. <u>Pre-differentiation of hE SCs using 1 00ng/ml activin A in s erum free conditions for 5-8 days into endoderm *in vitro*. Confirm endoderm phenotype using immunohistochemistry and FACs analysis.</u>

- This task was to be performed in collaboration with Professor Alan Trounson. Since the project began, Prof. Trounson has left academ ic research and now heads the Californian Institute Regenerative Medicine, CA, USA. Since his departure, we have established links with scientists in his laboratory to transfer this technology to our site at MIMR. This has b een relatively successful, but has taken som e time to optimize conditions of pre-differentiation in our hands. We can now reliably and reproducibly produce ~60-80% definitive e ndoderm from hESCs using activin A. As described in the proposal, we have used dual fluores cent labeling with Sox17 and CXCR4 to identify the purity of our cell populations. Attempts are currently being made to FACs sort out these cells to en rich for more pure populations that will lim it the spontaneous differentiation of hESCs when we use them in tissue recom bination assays.
- c. <u>Generation of tissue recom</u> <u>binants of endode rm-derived hESCs together with rodent UGM or SVM (isolated from E17.5 m</u> <u>ale e mbryos for UGM or day 0 m</u> <u>ale pups for SVM) using collagen gel technique and sub-renal grafting into male SCID mice.</u>
  - We recently initiated tissue recombination studies, following ethics approval that was granted in December 2008. This is an established protocol that is routinely performed in our laboratory. At present, we have set up 2 series of tissue r ecombination experiments that utilized undifferentiated hESCs and pre-differentiated hESCs. The grafts are presently growing in host male mice and tissues are scheduled to be harvested in April 2009.
- d. <u>Harvesting and analysis of tissue recom</u> <u>binants including immunohistochem istry for morphological analysis and cell death/proliferation markers.</u>
  - No progress to date.
- e. <u>ALTERNATIVE METHOD</u>: perform two-step tissue r ecombination with endoderm derived hES Cs and rodent UGM or SVM us ing collagen gel technique and sub-renal grafting into male SCID mice, if first method is not optimal.
  - No progress to date.

### **Key Research Accomplishments:**

- Establishment and maintenance of hESC cultures.
- Performed pre-differentiation of hESCs into definitive endoderm.
- Initiate tissue recombination studies with undifferentiated and pre-differentiated hESCs (yet to be analysed).

### **Reportable Outcomes:**

- Manuscripts:
- 1. Taylor RA, Risbridger GP (2008) The path towards identifying prostatic stem c ells. **Differentiation** 76(6):671-681 (*IF 3.745*) *JRank 53/156 Cell Biology*

- 2. Taylor RA, Risbridger GP (2008) Prostatic tumour strom a: a key player in cancer progression. Current Cancer Drug Targets 8(6):490-7 (IF 5.677) JRank 17/127 Oncology
- 3. Risbridger GP, Taylor RA (2008) Prostatic stem cell niche in health and disease. **Endocrinology** 149(9):4303-4306 (*IF* 5.236) *JRank* 13/93 Endocrinology & Metabolism

# Abstract presentations:

1. Taylor RA, Toivanen R, Pedersen J, Collins A, Maitland NJ, Risbridger GP (2008) Altered differentiation of CD133+ prostatic stem cells by carcinoma-associated fibroblasts. *The Role of Cancer Stem Cells in the Initiation and Propagation of Tumorigenesis; Special conference of American Association for Cancer Research*, Los Angeles, USA (*poster presentation*).

## **Conclusion:**

In summary, work towards task 1 of this project has begun and it is anticipated that we will begin work towards task 2 in the next 2-4 months. Task 2 involves the isolation of prostatic stem cells from our hESC-derived tissues. Recently there have been several publications that will enable us to isolate enriched prostatic stem cell population using newly identified cell surface makers including CD117 (Leong, et al., 2008) or alternative markers such as Trop2 (Goldstein, et al., 2008). These advances will lead to reliable isolation of prostatic stem cells that have proven regenerative capacity with which to test the effects of prostatic tumor stroma.

The long term goal of defining the role of pr ostatic tum our strom a in the initiation of carcinogenesis will have a great impact on the field of prostate cancer (and other major cancers) leading to funda mental changes in our thin king about cancer therapy. If the outcom es demonstrate that stroma plays a key role in the in itial stages of tumorigenesis, further work will be required to define the strom ally-derived factors that may become novel target molecules for early stage prostate cancers.

### References

Taylor RA, Cowin PA, Cunha GR, Pera M, Trounson AO, Pedersen J and Risbridger GP (2006) Formation of human prostate tissue from embryonic stem cells. Nat Methods 3:179-181 Leong KG, Wang BE, Johnson L and Gao WQ (2008) Generation of a prostate from a single adult stem cell. Nature 456:804-808

Goldstein AS, Lawson DA, Cheng D, Sun W, Garraway IP and Witte ON (2008) Trop2 identifies a subpopulation of murine and human prostate basal cells with stem cell characteristics. Proc Natl Acad Sci U S A 105:20882-20887

# **Appendices**

N/A